MicroRNA-7-5p Inhibits Migration, Invasion and Metastasis of Intrahepatic Cholangiocarcinoma by Inhibiting MyD88

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Abstract

Background and Aims: Intrahepatic cholangiocarcinoma (ICC) is a malignant tumor derived from intrahepatic bile duct epithelial cells. Accumulating studies report that microRNAs are widely involved in tumor migration and metastasis by regulation of target genes. miR-7-5p has been confirmed to inhibit tumor metastasis and to be related to prognosis for several malignant tumors. Our study investigated the underlying functions of miR-7-5p in ICC.

Methods: The expression of miR-7-5p in ICC tissues but also in ICC cell lines was analyzed by real-time PCR. By analyzing the relationship between the clinicopathological parameters of 60 ICC patients and the expression level of miR-7-5p, the effect of miR-7-5p on the prognosis was clarified. After transfected with miR-7-5p mimics or miR-7-5p inhibitor, cell counting kit-8 assay was applied to evaluate the cells proliferation, flow cytometry was applied to analyze the cells apoptosis, wound healing assay and transwell chamber assay were applied to analyze the cell invasion and migration. A luciferase reporter assay was identified the relationship of miR-7-5p and myeloid differentiation factor 88 (MyD88). Western blotting was used to analyze the proteins expression. And immunohistochemistry was performed to determine the expression of MYD88 in ICC tissues. Results: Our data showed the expression of miR-7-5p was down-regulated not only in ICC tissues but also in ICC cell lines compared with normal controls. Low expression of miR-7-5p was notably associated with poor prognosis in ICC patients. miR-7-5p negatively regulated cell proliferation, migration, invasion and apoptosis in ICC cells. We further verified that MyD88 was a novel target of miR-7-5p and was significantly overexpressed in ICC tissues. Overexpression of MyD88 counteracted the effects of miR-7-5p in ICC cells. Conclusions: The present findings suggest that miR-7-5p plays a pivotal role in ICC invasion by regulating MyD88. Ample evidence into the key factors of ICC invasion may result in the development of new treatment options for ICC.

Keywords: Intrahepatic cholangiocarcinoma; miR-7-5p; MyD88.

Abbreviations: ANTs, adjacent non-tumor tissue samples; CCK-8, cell counting kit-8; EMT, epithelial-mesenchymal transition; HCC, hepatocellular carcinoma; ICC, intrahepatic cholangiocarcinoma; IL-1R, interleukin 1 receptor; miRNAs, microRNAs; MyD88, myeloid differentiation factor 88; T-ALL, T cell acute lymphoblastic leukemia; TLR, toll-like receptor; UTR, untranslated region.

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Introduction

Intrahepatic cholangiocarcinoma (ICC) is a very aggressive malignant tumor of the biliary tract, which is often diagnosed in the advanced stage.1-2 Surgery is the main curative option. First-line chemotherapy is the preferred strategy in cases with macroscopic residual disease after surgery or locally recurrent or unresectable cholangiocarcinoma at the time of diagnosis.3 However, the treatment effect is very poor. The long-term survival time of patients with unresectable ICC is very short, and the 5-year survival rate after diagnosis is less than 5% to 10%.3,4 Therefore, it has important clinical significance for finding new prognostic indicators and therapeutic targets by exploring the molecular mechanism of the progression of ICC.5-7 MicroRNAs (miRNAs) are highly conserved small noncoding RNAs, with lengths of approximately 17–25 nucleotides. Recently, miRNAs have been shown to be involved in multiple steps that regulate the occurrence and development of ICC.8,9 MI-7-5p has been confirmed as exerting antitumor effects in multiple types of carcinomas, including breast cancer, pancreatic ductal adenocarcinoma, glioma, etc.9-11 Therefore, the underlying mechanisms and significance of miR-7-5p in ICC remain unclear.

Myeloid differentiation factor 88 (MyD88) mainly mediates the inflammatory response to cell injury as an adapter protein binding to the Toll-like receptor (TLR) and interleukin 1 receptor (IL-1R).12-14 Previous studies have shown that the abnormal expression of MyD88 is closely related to tumor development and drug resistance. MyD88 is abnormally highly expressed in many kinds of malignant tumor cells and tissues.12,13,16 However, there are other research data showing that MyD88 exerts an anti-tumor effect. Therefore, there is still no conclusion about the role of MyD88 in the occurrence and development of tumors.12,13,15,17

In this study, we studied the expression level of miR-7-5p in ICC tissues and cells and its clinicopathological relationship with ICC cases, and explored the possible mechanism of miR-7-5p inhibiting the migration and invasion of ICC cells.
Methods

Surgical specimens and clinicopathological data

From January 2013 to January 2018, primary cancer tissues with pathologically confirmed ICC (n=60) and their matched adjacent non-tumor tissue samples (ANTs) were obtained at the People's Hospital of Guangxi Zhuang Autonomous Region. The clinical characteristics of the ICC patients are described in Table 1. After surgery, the ICC patients were followed up for 1–70 months. Patients' disease-free survival time was determined from the operation date to the date of recurrent disease diagnosis or the end of follow-up. Overall survival time was determined from the operation date to the date of death or the end of follow-up. All tissues were stored at −80°C prior to use. All tissues were acquired with written informed consent. Our research plans were approved by the Clinical Research Ethics Committee of the People’s Hospital of Guangxi Zhuang Autonomous Region (Number: 2013-09), according to the Helsinki Declaration.

Cell culture and transfection

Our study used several ICC cell lines, of which HCCC-9810, HuCCT1, QBC-939, and RBE were purchased from the Shanghai Cell Bank, Chinese Academy of Sciences (Shanghai, China). The human primary intrahepatic biliary epithelial cell line HIBEC was purchased from ATCC (Manassas, VA, USA). All cells were cultured in DMEM (Thermo Fisher, Waltham, MA, USA) according to the manufacturer’s recommendations. Hsa-miR-7-5p mimics (miR1115129142345-1-5), hsa-miR-7-5p inhibitor (miR2170523092350-1-5) and negative control vector were obtained from RiboBio (Guangzhou, China). MyD88 lentiviral activation particles (sc-417166-ACT) and control lentiviral activation particles (sc-437282) were purchased from Santa Cruz Biotechnology (Dallas, TX, USA). RBE cells were transfected with miR-7-5p mimics or negative control vector, co-transfected with miR-7-5p mimics and MyD88 lentiviral activation particles or control lentiviral activation particles. HIBEC cells were transfected with negative control vector or hsa-miR-7-5p inhibitor. Cells were transfected 24 h before experiments.

Cell counting kit-8 (CCK-8) assay

Two groups of RBE cells were digested with trypsin and seeded into 96-well plates. Approximately 400,000 cells were added to each well. Then, the cells were cultured in the incubator for 24, 48 or 72 h. Next, the cells were exposed to CCK-8 reagent, mixed, and incubated for 1 h. An enzyme-linked immunosorbent detector (Dojindo Molecular Technologies, Kumamoto, Japan) was employed to estimate the absorbance at 450 nm to generate a growth curve.

Flow cytometric analysis of apoptosis

RBE cells were harvested after 48 h, and apoptotic rates were detected by Annexin-V/FITC Kit (BD Biosciences, San Jose, CA, USA). In short, RBE cells were washed with pre-chilled phosphate-buffered saline, added to 1 mL of 1× Annexin V-FITC binding buffer, centrifuged, and (pellet) resuspended by adding 200 µL binding buffer. RBE cells were then added to 10 µL Annexin V-FITC and 5 µL propidium iodide and incubated for 30 m in the dark. Finally, the apoptotic rate of each group was detected by a FACSCalibur flow cytometer (BD Biosciences). The log of FITC-annexin

Table 1. Correlation of mir-7-5p expression with clinicopathological characteristics in intrahepatic cholangiocarcinoma

<table>
<thead>
<tr>
<th>Variables</th>
<th>Cases, n=60</th>
<th>miR-7-5p expression</th>
<th>p</th>
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<tbody>
<tr>
<td></td>
<td>Low, n=30</td>
<td>High, n=30</td>
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<tr>
<td>Age in years</td>
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</tr>
<tr>
<td>≤55</td>
<td>25</td>
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<td>14</td>
</tr>
<tr>
<td>&gt;55</td>
<td>35</td>
<td>19</td>
<td>16</td>
</tr>
<tr>
<td>Gender</td>
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</tr>
<tr>
<td>Female</td>
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<td>9</td>
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</tr>
<tr>
<td>Tumor stage</td>
<td></td>
<td></td>
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<tr>
<td>T1+T2</td>
<td>40</td>
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<td>24</td>
</tr>
<tr>
<td>T3+T4</td>
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<td>6</td>
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</tr>
<tr>
<td>Negative</td>
<td>44</td>
<td>18</td>
<td>26</td>
</tr>
<tr>
<td>Positive</td>
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<td>12</td>
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<tr>
<td>High</td>
<td>30</td>
<td>22</td>
<td>8</td>
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</table>

*Statistically significant.
V–fluorescence was displayed on the x-axis, and the log of propidium iodide fluorescence was displayed on the y-axis. For each analysis, 10,000 events were recorded.

Transwell chamber assay

Matrigel (BD Biosciences) was added in the upper chamber. RBE cells were trypsinized and resuspended in serum-free medium. Approximately 40,000 cells were added to the upper layer, and 500 µL of Dulbecco’s modified Eagle’s medium was supplied to the lower layer. After phosphate-buffered saline wash, the noninvasive cells in the upper layer were gently wiped with a wet cotton swab, and the remaining cells were fixed with 4% paraformaldehyde for 20 min. After washing 3 times with phosphate-buffered saline, the cells were stained with 0.1% methyl violet for 25 min. After washing, five fields under the microscope were randomly selected to count the stained cells.

Wound healing assay

The transfected RBE cells or HIBEC cells were seeded into 6-well plates at the density of approximately 500,000. After the cells adhered to 90% confluence, they were uniformly scratched with a 200 µL pipette tip. Then, the cells were observed under a microscope and photographed after the floating cells were removed. After 24 h, they were observed again, and the migration areas were measured in five random fields.

Database analysis

We used the online target gene prediction databases of TargetScan 7.2 (http://www.targetscan.org/vert_72/) and MicroT (https://mir.tarbase.mbc.nctu.edu.tw) to jointly predict the miR-7-5p target genes, and then the two results were intersected. In addition, target gene databases with experimental verification support, miRTarBase (http://mir.tarbase.mbc.nctu.edu.tw) and miRWalk (http://129.206.7.150/), were jointly used to detect possible targets of miR-7-5p. The results were combined as the final miR-7-5p target gene prediction result.

Dual-luciferase reporter gene assay

A luciferase reporter assay with the MyD88 3′-untranslated region (UTR) was used to verify that MyD88 was the direct target of miR-7-5p. The negative control vector or miR-7-5p mimics were co-transfected with the constructed WT-MyD88-3′-UTR (wild-type seed sequence: CUGCCCU) or MT-MyD88-3′-UTR (mutant seed sequence: CUAGGGU) psiCHECK2 reporter vector (Promega, Madison, WI, USA) into RBE cells using Lipofectamine 2000 according to the manufacturer’s instructions. Renilla and firefly luciferase activities were assessed with a dual-luciferase reporter gene assay system (Promega).

Western blotting

Total protein were extracted separately from ICC cells. A separation gel and concentrated gel were prepared, and the proteins were separated by electrophoresis and then transferred to membranes. The membranes were blocked with 5% skim milk and incubated with antibodies against MyD88 (1:1,000; Santa Cruz Biotechnology), IRAK4 (1:1,000), TRAF6 (1:1,000), NF-κB (1:1,000), phosphorylated (p)-NF-κB (1:1,000), GAPDH (1:2,000; Proteintechn, Hubel, China) and washed after incubation. Then, the membranes were incubated with goat anti-rabbit/mouse secondary antibody (1:3,000; Multisciences, Hangzhou, China) at room temperature for 1 h. After being washed, the poly (vinylidene difluoride) membrane was placed in enhanced chemiluminescence reagent and then scanned.

RNA extraction and real-time PCR analysis

Total RNA was extracted from ICC frozen tissues and ICC cell lines by the TRIZol method (Invitrogen, Carlsbad, CA, USA) and stored in a freezer at −80°C. Then, the RNA was reverse transcribed to synthesize complementary DNA by using the miRNA qPCR Detection kit (GeneCopoeia, Rockville, MD, USA) according to the kit instructions. The primer sequences used for quantitative reverse transcription-PCR were as follows: 5′-AAAACCTGTGCGCAAACCA C-3′ (forward) and 5′-GCTGCATTATACGAGCCAAGTAAG-3′ (reverse) for miR-7-5p; 5′-GGGTCTCTCCACATGGA-3′ (forward) and 5′-CTGTGCTCCGAGTTCAAGA-3′ (reverse) for MyD88; and 5′-CTGCCTCTCCAGCAAGCACA-3′ (forward) and 5′-AACCCTTCAGGATATG-3′ (reverse) for U6. MyD88 data were normalized to GAPDH, which was detected using the following primers: 5′-GAACACCACAACTCGTCTAGCA-3′ (forward) and 5′-TCTCTGCGGCTGCACTTGGTG-3′ (reverse), while U6 was used as an endogenous control for miR-7-5p.

Immunohistochemistry staining of MyD88

The formalin-fixed and paraffin-embedded ICC tissues were cut into 4-µm thick sections, and then we performed antigen retrieval and suppressed endogenous peroxidase activity. MyD88 (1:300; Santa Cruz Biotechnology) was used as the primary antibody to incubate the slides at 4°C overnight, which was followed by incubation with a secondary antibody (1:500; MultiSciences). Finally, the DAB solution was applied to signal generation, and the slides were stained again with hematoxylin and observed by an optical microscope.

Statistical analysis

Data expressed as the mean±standard error of the mean were analyzed by SPSS 22.0 software (IBM Corp., Armonk, NY, USA). Experiments were performed at least three times. We used the chi-square test to explore the correlation between the two variables. For continuous variable analysis, independent t-tests and variance A-tests were used. The overall survival and disease-free survival rates of the patients were measured by the Kaplan–Meier method, and the survival rate between the groups were compared by a log-rank test. Prognostic factors were analyzed by univariate analysis and multiple Cox regression modeling. Correlation curves were constructed, and differences between groups were calculated by using Spearman rank correlation analysis. A p-value less than 0.05 was considered to indicate a statistically significant difference.

Results

miR-7-5p was down-regulated in ICC tissues and cells

In previous reports, miR-7-5p was often negatively regu-
lated in ICC tissues. To further verify this observation, we first measured the expression of miR-7-5p (written as miR-7 hereafter for conciseness) in 60 pairs of ICC tissues and ANTs by real-time PCR. Notable down-regulation of miR-7 was observed in ICC tissues compared with the matched ANTs (Fig. 1A). To explore whether miR-7 was dysregulated in ICC cells, the expression of miR-7 in ICC cell lines (HCCC-9810, HuCCT1, QBC-939, RBE) and normal HIBEC cells were quantified. We confirmed that the expression of miR-7 was also reduced in ICC cell lines compared with HIBEC cells (Fig. 1B). Therefore, we demonstrated the frequent down-regulation of miR-7 in ICC.

**Down-regulation of miR-7 was related to poor prognosis**

To reveal the clinicopathological characteristics of miR-7 in ICC, we divided ICC patients into two groups according to the median expression of miR-7: the miR-7 low expression group (n=30) and the miR-7 high expression group (n=30). The results showed that the low expression of miR-7 was related to higher tumor grade, more positive lymph node metastasis, and higher clinical stage (Table 1). Because poorer cellular differentiation and advanced tumor stage were related to ICC metastasis, the down-regulation of miR-7 may participate in the metastatic growth of ICC. In addition, we also investigated the relationship between low miR-7 expression and patients' survival. After 3 years of follow-up, we found that the 1-year disease-free survival rates and 3-year overall survival rates of patients in the low miR-7 expression group were notably poorer than those in the high expression group (Fig. 1C, D). All the above data revealed that the loss of miR-7 expression may play a role in the progression of ICC and causing the poor prognosis of ICC patients.

**miR-7 negatively regulated RBE cell proliferation, migration, and invasion and induced apoptosis**

We observed that the deletion of miR-7 was closely correlated with the metastasis and prognosis of ICC; further, we analyzed the effect of miR-7 on ICC cell proliferation, migration, and invasion. Because miR-7 had the lowest expression in RBE cells, we first constructed a stable expression of miR-7. Compared with the negative control group, miR-7 expression was confirmed to notably increase in the miR-7 mimic transfection group (Fig. 2A). Through the CCK-8 assay, we observed that miR-7 re-expression notably inhibited RBE cell proliferation in vitro (Fig. 2B). We then used flow cytometry to reveal the effect of miR-7 re-expression on apoptotic rates. The data indicated that the apoptotic rate of RBE cells also increased markedly with increasing miR-7 expression, which demonstrated that miR-7 induced cell apoptosis in ICC (Fig. 2C). In the Transwell cell invasion assay, overexpression of miR-7 reduced the cells which invaded the lower chamber (Fig. 2D). Similarly, in the miR-7 high expression group, the ability of cells to migrate was notably impeded compared with that in the low expression group (Fig. 2E). To further detect the effects of miR-7 on cell migration and invasive ability, HIBEC cells expressing relatively high endogenous miR-7 were transfected with miR-7 inhibitors. The down-regulation of miR-7 was verified by quantitative reverse transcription-PCR (Fig. 2F). Interestingly, inhibition of miR-7 markedly promoted cell invasion (Fig. 2G) and migration in HIBEC cells (Fig. 2H). Our prior studies demonstrated that miR-7 suppressed ICC metasta-
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**MyD88 was a direct target of miR-7 in RBE cells**

Previous research results revealed that miR-7 functions as a tumor suppressor gene in ICC. Accordingly, we further sought to identify the underlying targets of miR-7 that were related to tumor metastasis. We explored miR-7 downstream gene targets through four different miRNA targeting prediction tools: TargetScan, MiRanda, miRTarBase and miRwalk. A potential candidate target gene, MyD88, was then selected, which had a predicted consequential pairing with miR-7 at bases 55–61 (Fig. 3A). To further explore whether the anti-metastatic properties of miR-7 in ICC were related to MyD88, we first performed a dual-luciferase reporter gene assay in RBE cells. The luciferase activity of RBE cells with wild-type MyD88 after transfection with miR-7 mimics was markedly reduced compared with that of the mimic NC-transfected cells. However, in RBE cells with mutated MyD88, the decrease in luciferase activity was significantly reversed (Fig. 3B). We further verified whether miR-7 could regulate MyD88 expression at the cellular level.

After transfection with miR-7 mimics, the MyD88 level of RBE cells was decreased by approximately 50% (Fig. 3C). The inhibition of miR-7 in normal HIBEC cells markedly increased MyD88 expression (Fig. 3D). Accordingly, MyD88 was proven to be a novel target of miR-7 in RBE cells.

**Overexpression of MyD88 counteracted the effects of miR-7 in RBE cells**

Previous findings indicated that miR-7 inhibits metastasis by negatively regulating ICC cell migration and invasion and that MyD88 was directly targeted by miR-7. Therefore, we suspected that MyD88 may participate in miR-7-mediated anti-metastatic properties. To verify that miR-7 directly targets MyD88 to perform its function, miR-7-transfected RBE cells were infected with MyD88 lentiviral activation particles or control lentiviral activation particles. MyD88 protein levels were significantly increased by transfection of MyD88 lentiviral activation particles (Fig. 4A), and mRNA levels exhibited the same trend with quantitative reverse transcription-PCR detection (Fig. 4B). In wound healing assays, the recovery of MyD88 expression partially promoted the migration of RBE cells overexpressing miR-7 (Fig.
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4C). In Transwell assays, increased expression of MyD88 enhanced the invasion ability of miR-7-overexpressing RBE cells, which was suppressed by miR-7 (Fig. 4D). Accordingly, the findings showed that miR-7 inhibited migration and invasion by targeting MyD88, and the underlying mechanism was further studied. The IRAK4/TRAF6/NF-κB signaling pathways, which had been reported to be important pathways for regulating the differentiation, survival, and movement of normal and cancer cells, were closely related to the physiological role of MyD88. Therefore, we further investigated the influences of miR-7 and MyD88 on the expression changes of IRAK4, TRAF6, NF-κB and p-NF-κB. Western blotting analysis revealed that overexpression of miR-7 suppressed IRAK4 and TRAF6 protein expression and negatively regulated NF-κB phosphorylation. The restoration of MyD88 expression in RBE cells notably reversed the inhibitory effects of miR-7 (Fig. 4E). All these results suggested that miR-7 inhibited invasion and metastasis by directly targeting MyD88, possibly through IRAK4/TRAF6/NF-κB/MyD88 signaling pathways.

MyD88 was significantly overexpressed in ICC and negatively correlated with the expression of miR-7

Due to the promoting effect of MyD88 in many tumors, we first determined the expression of MyD88 in ICC tissues. ICC tissues showed significantly higher expression of MyD88 than ANTs (Fig. 5A). Interestingly, we also found a negative correlation between miR-7 and MyD88 expression (Fig. 5B). Furthermore, we examined the expression of MyD88 in ICC cell lines. The expression level of MyD88 was confirmed to be increased in all ICC cell lines through quantitative reverse transcription-PCR, with the highest expression in RBE and QBC-939 cells (Fig. 5C). We further quantified the expression levels of MyD88 in ICC tissues with different miR-7 levels by immunohistochemistry (Fig. 5D). Tumors with lower levels of miR-7 tended to overexpress MyD88, and conversely, tumors that overexpressed miR-7 showed a lower level of MyD88. After 3 years of follow-up, we found that the 1-year disease-free survival rates and 3-year overall survival rates of patients in the high MyD88 expression group were notably poorer than those in the low expression group (Fig. 5E, F). Univariate Cox proportional hazard regression analyses showed that tumor-necrosis-metastasis (known as TNM) stage, miR-7-5p expression, and MyD88 expression were prognostic factors of ICC (p<0.05). See Table 2 for details. Including TNM tumor stage, miR-7-5p expression, and MyD88 expression into the Cox model and performing multivariate analysis showed that MyD88 expression (odds ratio: 3.834, 95% confidence interval: 1.435–10.239, p<0.05) was an independent factor affecting prognosis. Collectively, these data suggested that dysregulation of miR-7 was related to the overexpression of MyD88 in ICC and high MyD88 expression was related to poor prognosis of ICC.

Discussion

ICC is a malignant tumor derived from the bile duct epithelium and ranks second among the primary liver cancers.1,18 The difference between ICC and extrahepatic cholangiocarcinoma is that ICC occurs in the liver parenchyma. Although the incidence of ICC worldwide is much lower than that of hepatocellular carcinoma (HCC), some recent studies have
The only potentially curative treatment for ICC is radical surgery. However, less than 30% of patients undergo surgical resection because the early diagnosis rate of ICC is very low. Moreover, the 5-year survival rate after ICC surgical resection is less than 45%, and the recurrence rate is about 50% from most clinical studies. Most patients have lost the opportunity for radical surgery. For patients with advanced unresectable and metastatic ICC, the standard first-line systemic treatment is gemcitabine combined with cisplatin. But the therapeutic effect is very limited, and the overall survival time is less than 1 year. New effective treatment methods are urgently required for ICC patients.

MiR-7 plays a role in the diagnosis, prognosis, anti-metastasis and treatment of various cancers via targeting different genes in various complex ways. MiR-7 can induce rhabdomyosarcoma cell necrosis and apoptosis via mitochondrial damage. MiR-7-5p also can modulate malignant phenotypes of T cell acute lymphoblastic leukemia (referred to as T-ALL) cells and serve as a potential therapeutic target for T-ALL. Moreover, miR-7 acts as a tumor suppressor in digestive system malignancies. MiR-7 is down-regulated in HCC, which directly targets the tumor suppressor gene CUL5 towards involvement in the progression of HCC. MiR-7 is also down-regulated in colorectal cancer to negatively regulate the tumorigenesis gene PAX6 towards involvement in the progression of colorectal cancer. These studies show that miR-7 can play an anti-tumor role in digestive system tumors by acting on various key genes.

ICC are clinically characterized by poor prognosis, partly due to tumor angiogenesis via VEGF overexpression. In liver cancer, it has been found that over-activated TLR4/MyD88 can up-regulate the expression of VEGF and participate in tumor progression in HCC. But the role of MyD88 in ICC is temporarily poorly understood. Meanwhile, it is reported that the MyD88 signaling could play dual functional roles in colorectal cancer. Sometimes, it plays the tumor-promoting role that enhances cancer inflammation and intestinal flora imbalance to induce tumor invasion and tumor cell self-renewal; at other times, it plays an anti-tumor role that helps to maintain the host-microbiota homeostasis to induce tumor cell cycle arrest and immune responses against cancer cells. Thus, studying the role of MyD88 in cholangiocarcinoma is not only interesting but also meaningful. More and more reports indicate that MyD88 combined with NF-κB promotes tumor development. NF-κB is one of the important signaling molecules downstream of MyD88. Notably, we confirmed that MyD88 was not only increased in many ICC cell lines but also significantly higher in ICC tissues than in ANTs. MyD88 was then predicted to be a downstream gene target of miR-7 by four different
miRNA targeting prediction tools, and the targeting relationship was confirmed through a dual-luciferase reporter gene assay. Interestingly, the present data revealed that miR-7 was negatively correlated with MyD88 in ICC tissues. Tumors with lower levels of miR-7 tended to overexpress MyD88, and conversely, tumors that overexpressed miR-7 showed a lower level of MyD88. MyD88 expression was an independent factor affecting prognosis. Furthermore, restoration of MyD88 rescued miR-7-inhibited tumor migration and invasion. All the findings showed that miR-7 inhibits tumor metastasis by targeting MyD88 in vitro.

Previous studies have shown that MyD88 is required for IRAK4/TRA66/NF-κB signaling, which plays a pivotal role in the phenotypic changes, invasion and metastasis of malignant tumor cells by promoting epithelial-mesenchymal transition (commonly referred to as EMT) and local adhesion dynamics. Overexpression of MyD88 is involved in activating NF-κB, inducing cell proliferation, anti-apoptosis, and promoting cell EMT. With increased p-NF-κB expression levels, cell migration and invasion capabilities are enhanced.

Therefore, we further detected the influences of miR-7 and MyD88 on the expression changes of IRAK4, TRAF6, NF-κB and p-NF-κB. We confirmed that up-regulation of miR-7 inhibited IRAK4 and TRAF6 protein expression and negatively regulated the phosphorylation of NF-κB. The restoration of MyD88 in RBE cells notably reversed inhibitory effects of miR-7. Therefore, we postulated that miR-7 could inhibit invasion and metastasis in ICC by directly targeting MyD88, possibly through IRAK4/TRA66/MyD88 signaling pathways. Further research investigations are needed to adequately illuminate the miR-7/MyD88-related signaling pathway and its functions in ICC.

miR-7 was confirmed to significantly inhibit metastasis by down-regulating MyD88 in vitro. Similarly, Shen et al. demonstrated that miR-7 inhibited invasion in colorectal cancer by regulating the twist family bHLH transcription factors (TGF-β).
factor 1 and the down-regulation of miR-7, promoting lung metastasis. The prior studies showed that miR-7 could act as a tumor suppressor in many types of malignancies.

In summary, we confirmed the down-regulation of miR-7-5p in ICC, which was related to poor prognosis. miR-7-5p inhibited invasion and metastasis in vitro by directly targeting MyD88, indicating that miR-7-5p and MyD88 could be novel targets for preventing ICC metastasis in the future.

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None to declare.

**Conflict of interest**

The authors have no conflict of interests related to this publication.

**Author contributions**

Conception and design of the project (YtT, ZT), performance of the experiments (ZW, CC, XY), writing of the manuscript (TL), critical revision of the manuscript (YtT, ZT). All authors read and approved the final manuscript.

**Ethics approval and consent to participate**

All tissues used in this study were acquired with written informed consent. All human experiments were approved by the Clinical Research Ethics Committee of the People’s Hospital of Guangxi Zhuang Autonomous Region (No. 2013-09). All protocols involving the use of animals were approved by the Animal Experimental Ethical Inspection of the People’s Hospital of Guangxi Zhuang Autonomous Region.

**Data sharing statement**

The datasets used and analyzed during the current study are available from the corresponding author, upon reasonable request.

**References**


